	<h1 style="margin: 0;">THE GOODELL LABORATORY</h1>																			
Author	Nathan Boles	Feb.5, 2009																		
Title	Peripheral Blood Analysis																			
Introduction	This protocol describes the analysis of mouse peripheral blood																			
Materials	<p>HBSS+ : Hanks Balanced Salt Solution (from Gibco) with 2% Fetal Calf Serum and 10 mM 2% dextran in PBS Heparin Solution Antibody Cocktail (In HBBS+)</p> <table border="0" style="width: 100%; border-collapse: collapse;"> <thead> <tr> <th style="text-align: left; border-bottom: 1px solid black;"><u>Antibody</u></th> <th style="text-align: right; border-bottom: 1px solid black;"><u>Concentration</u></th> </tr> </thead> <tbody> <tr><td>B220 - Pac Blue</td><td style="text-align: right;">1:100</td></tr> <tr><td>B220 - Pe-Cy7</td><td style="text-align: right;">1:100</td></tr> <tr><td>CD4 - Pac Blue</td><td style="text-align: right;">1:100</td></tr> <tr><td>CD8 - Pac Blue</td><td style="text-align: right;">1:100</td></tr> <tr><td>Gr-1 - Pe-Cy7</td><td style="text-align: right;">1:200</td></tr> <tr><td>Mac-1 – Pe-Cy7</td><td style="text-align: right;">1:200</td></tr> <tr><td>CD45.1 - Fitc</td><td style="text-align: right;">1:100</td></tr> <tr><td>CD45.2 – APC</td><td style="text-align: right;">1:100</td></tr> </tbody> </table>		<u>Antibody</u>	<u>Concentration</u>	B220 - Pac Blue	1:100	B220 - Pe-Cy7	1:100	CD4 - Pac Blue	1:100	CD8 - Pac Blue	1:100	Gr-1 - Pe-Cy7	1:200	Mac-1 – Pe-Cy7	1:200	CD45.1 - Fitc	1:100	CD45.2 – APC	1:100
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Protocol		<i>Notes</i>																		
	<ol style="list-style-type: none"> 1. Prepare 1.5 mL tubes to receive blood by pipeting 150μL of 2% dextran in PBS and 150μL of Heparin Solution. 																			
	<ol style="list-style-type: none"> 2. Retro-orbitally bleed mice using heparin coated capillary tubes and place blood into prepared 1.5mL tubes. Also obtain splenocytes to use as a control at this time. 																			
	<ol style="list-style-type: none"> 3. Allow blood to settle for at least 20 minutes (until supernatant appears almost white) then pipet off supernatant into an appropriate FACS tube. 																			
	<ol style="list-style-type: none"> 4. Add 1 mL of RBC lysis buffer to each tube. 	...																		

5	Wait at least ten minutes.	<i>Best to wait around 20 minutes</i>
6	While you are waiting, prepare the antibody cocktail shown in the Materials section of this protocol(enough to make a final solution of 100 μ L per sample).	<i>CD45.1 or CD45.2 can be substituted for an antibody of your choice. Also PE is still open for the use of an antibody of your choice.</i>
7	Place antibodies into an appropriate amount of Hank's+ (enough to make 100 μ L per tube) to finish your antibody cocktail.	
8	Stain spleenocytes with B220+ in each color to act as controls	
9	Wash tubes with 2 mL of Hank's+, then spin down for 8 min. at 2000 rpm.	
10	Discard supernatant (I personally use the 'net method' in combo with the centrifuge holders)	<i>In order to discard supernatant of all the tubes together, you can use a plastic web to cover the tubes and turn it down together with the centrifuge holders.</i>
11	Resuspend cells with 100 μ L of the antibody cocktail	
12	Wait 10 minutes, then wash with 2 mL of Hank's+ and spin down for 8 min. at 2000 rpm.	
13	While your samples are spinning prepare a PI solution by using the lab stock of PI (1:100concentration) and Hank's+.	
14	Discard supernatant.	
15	Resuspend cells with 300 μ L of PI solution.	
16	Go do the analysis on the LSRII.	

References.

1. Challen G, Boles NC, Lin KY, Goodell MA. Mouse Hematopoietic Stem Cell Identification And Analysis. *Cytometry A*. 2009 Jan;75(1):14-24.

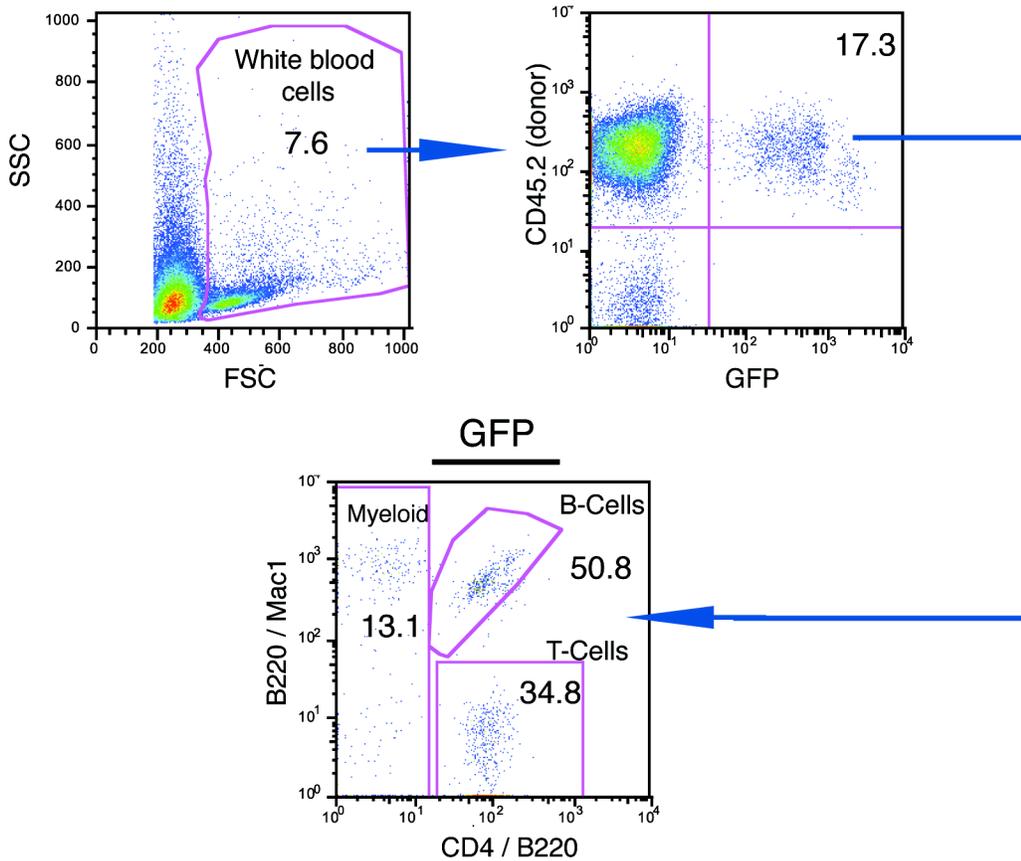


Figure 1. Example of Retroviral transduction and lineage analysis. At twelve weeks after transplant/transduction mice were bled and engrafted/transduced cells were identified using CD45.2-APC (donor background) and eGFP expression. B-cells were dual stained with B220-Pacific Blue and B220-PE-Cy7. T-cells were single labelled with CD4- and CD8-Pacific Blue. Myeloid cells were Pacific Blue negative. Transduced and non-transduced cells were simultaneously examined using a FacsAria (BD).