



Baylor College of Medicine Microarray Core Facility

One Baylor Plaza, Room 452A, Houston, Texas 77030
(713) 798-7699 microarray@bcm.tmc.edu



RNA Isolation (TRIZOL[®] Method)

1. Add tissue into 5 ml of TRIZOL[®] reagent to each tube and homogenize 6X for 10 seconds each (1 ml of TRIZOL[®] per 50-100 mg of tissue). Clean rotor with dH₂O twice and TRIZOL[®] reagent once between each sample.
2. Leave at room temperature for 5 minutes.
3. Add 1.0 ml of chloroform to each tube. Shake vigorously for 30 seconds (0.2 ml of chloroform per 1 ml of TRIZOL[®] reagent). Leave at room temperature for 3 minutes.
4. Separate phases by spinning at 10,000 rpm for 20 minutes without brake. Transfer the top phase into clean tubes (There should be about 1.6ml of top phase to take up).
5. Add 2 ml of isopropanol to each tube. Mix by inverting tubes and hold at room temperature for 10 minutes.
6. Pellet at 10,000 rpm for 20 minutes. Wash with 4 ml of 70% ethanol.
7. Air dry pellet and remove all aqueous phase. Recover RNA in DEPC water (or other suitable solution) and transfer into microcentrifuge tubes. Store samples at -80°C.

NOTE: Keep all reagents associated with RNA Isolation separate from other reagents to prevent degradation and contamination of RNA. Make sure you use sterile techniques whenever handling these reagents.